

INTERNATIONAL PRELIMINARY EXAMINATION REPORT
(PCT Article 36 and Rule 70)



Applicant's or agent's file reference RE/PG4938	FOR FURTHER ACTION See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/GB 03/03703	International filing date (day/month/year) 28.08.2003	Priority date (day/month/year) 30.08.2002
International Patent Classification (IPC) or both national classification and IPC C07K14/54		
Applicant GLAXO GROUP LIMITED et al.		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.
2. This REPORT consists of a total of 8 sheets, including this cover sheet.
- ☒ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of 48 sheets.

3. This report contains indications relating to the following items:

- I ☒ Basis of the opinion
- II ☐ Priority
- III ☒ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- IV ☐ Lack of unity of invention
- V ☒ Reasoned statement under Rule 66.2(a)(ii) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- VI ☐ Certain documents cited
- VII ☐ Certain defects in the international application
- VIII ☐ Certain observations on the international application

Date of submission of the demand 04.03.2004	Date of completion of this report 21.10.2004
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**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/GB 03/03703

I. Basis of the report

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17))*):

Description, Pages

1-48 received on 21.11.2003 with letter of 19.11.2003

Claims, Numbers

1-38 as originally filed

Drawings, Sheets

1/34-34/34 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
☐ the language of publication of the international application (under Rule 48.3(b)).
☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
☐ filed together with the international application in computer readable form.
☒ furnished subsequently to this Authority in written form.
☒ furnished subsequently to this Authority in computer readable form.
☒ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
☒ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
☐ the claims, Nos.:
☐ the drawings, sheets:

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5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)).
(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

see separate sheet

III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

☐ the entire international application,

☒ claims Nos. 36-38 with respect to industrial applicability

because:

☒ the said international application, or the said claims Nos. 36-38 relate to the following subject matter which does not require an international preliminary examination (specify):

see separate sheet

☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

☐ no international search report has been established for the said claims Nos.

2. A meaningful international preliminary examination cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

☐ the written form has not been furnished or does not comply with the Standard.

☐ the computer readable form has not been furnished or does not comply with the Standard.

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes: Claims	1-3,5-11,18,21,22,34-38
	No: Claims	4,12-17,19,20,23-33
Inventive step (IS)	Yes: Claims	1-3,5-11,18,21,22,34,35
	No: Claims	4,12-17,19,20,23-33,36-38
Industrial applicability (IA)	Yes: Claims	1-35
	No: Claims	

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2. Citations and explanations

see separate sheet

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Re Item I.6

1. The application contains a replacement sequence listing filed with telefax received 19.12.2003.

Re Item III

2. Claims 36-38 relate to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT (methods for treatment of the human or animal body). Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(I) PCT) (see also Item V, "Industrial applicability", below).

Re Item V

3. Reference is made to the following prior art documents:

D1: WO 01/34645 A (THE GOVERNMENT OF THE UNITED STATES OF AMERICA) 17 May 2001 (2001-05-17)
D2: WO 01/62287 A (ADVANCED BIOTHERAPY) 30 August 2001 (2001-08-30)
D3: WO 99/51643 A (THE PENN STATE RESEARCH FOUNDATION) 14 October 1999 (1999-10-14)
D4: WO 00/65058 A (M & E BIOTECH) 2 November 2000 (2000-11-02)

Novelty, inventive step (Art. 33(2) and (3) PCT)

4. The invention relates to immunogens for the treatment of diseases as e.g. allergies. Such diseases may be treated by neutralising the biological effects of interleukin IL-13 by inducing the production of antibodies against IL-13.
The proposed immunogenic therefor comprises, in principle, two different compounds, i.e. an IL-13 element and one or more foreign T-cell epitopes (see claim 1). According to the description (page 5, "Summary of the Invention"), the IL-13 immunogen is modified to include foreign T-cell helper epitopes. Such a T-cell epitope is a peptide that is able to bind to an MHC-class II molecule; the peptide than stimulates T-cells (see description, page 12, paragraph 2). Epitopes used in the

proposed invention are for example from the Tetanus or Diphtheria toxin (other epitopes may be used, see description, page 13).

- 5.1 Having regard to the prior art documents D1 or D3 (considered independently), the subject-matter of claims 1-3, due to its broad formulation in general and generic terms does not appear to be novel (Article 33(2) PCT or to involve an inventive step (Article 33(3) PCT).

D1 discloses the use of mutated IL-13 molecules as antagonists for treating allergy diseases. It is referred to chimeric molecules comprising IL-13 and one or more effector molecules (see D1, e.g. claims 29 and 32; see page 9, lines 9-26); such effector molecules can be, for example, the Diphtheria toxin or other toxins. Certain of said effector molecules (e.g. Diphtheria toxin) are identical to the so-called T-cell epitopes involved in the present invention. Furthermore, the chimeric molecules of D1 may be used in diseases which are mediated by the activity of the interleukin IL-13 (see D1, page 6, lines 7-22 and page 93, Example 23) in the same way as in the presently proposed invention. Thus the combination of an IL-13 element with a T-cell epitope (termed effector molecule in D1) as presently broadly defined in claims 1-3 is considered to be embraced by the prior art disclosure of D1.

The combination of IL-13 and an effector molecule like Diphtheria toxin is also known from D3 which describes a mutagenized IL-13 based chimeric molecule that can deliver effector molecules to cells overexpressing IL-13 receptors like cancer cells (see for example claim 11 of D3).

Concerning D2, it also discloses the treatment of allergic conditions with allergy vaccines based on divers compounds such as interleukins; interferon or histamine and their receptors. However, it appears that the combination of IL-13 and a foreign T-cell epitope as claimed in the present invention is not disclosed in D2.

Concerning D4, it discloses a down regulation of IL-5 where the compounds involved are similar to those of the present invention, i.e. the interleukin is IL-5 (not IL-13) and the T-cell epitope (or effector molecule) may be from a Diphtheria or Tetanus toxoid epitope (see D4, for example claims 3 and 12).

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- 5.2 The above objections (item 5.1) also apply to the subject-matter of claims 5-11, 18, 21, 22, 34 and 35 since these claims do not contain any features allowing to clearly differentiate said subject-matter from the prior art D1 or D3 (at least when said claims refer back or relate to one of the claims objected to under novelty and inventive step).
- 5.3 The subject-matter of claims 4, 12-17, 19, 20 and 23-33 appears to be novel and to involve an inventive step since the more specific features of said subject-matter characterizing the IL-13 element and/or the foreign T-cell epitope do not appear to be disclosed in the prior art and do not appear to be derivable therefrom in an obvious way.
- 5.4 The subject-matter of the method claims 36-38 may be considered as formally new, but not as involving an inventive step. Insofar as the vaccine involved in said claims is known, the formulation of a method claim involving such a (known) vaccine is considered as a trivial formulation which is not the result of an inventive activity.

Definition of the invention (Art. 6 and Rule 6.3(a) PCT)

- 6.1 The subject-matter of the claims objected to under novelty and inventive step (items 5.1-5.2) does not meet the requirements of Art. 6 and Rule 6.3(a) PCT in that the matter for which protection is sought is not sufficiently defined.

The vague general or generic terms like "IL-13 element" or "foreign T-cell epitope" (see e.g. claim 1) do not help to clearly limit the scope of said claims. Such terms also do not allow to recognize novelty of the proposed invention and/or to determine the features of the invention which are needed for inventive activity, thus for neutralising the biological effects of interleukin 13 (as already mentioned under item 4 above). Since the neutralisation or down-regulation of IL-13 is well known (see the cited prior art), the proposed invention should also be based on an advantageous or unexpected effect over the prior art combinations of any IL-13 element and T-cell epitopes.

- 6.2 It is to be noted that in certain claims (e.g. claim 12) acknowledged under Article 33 PCT, the features defining the invention should also be more precise in order to meet the requirements of Article 6 PCT.

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- 6.3 The immunogen of the (independent) claim 33 should be defined per se in this claim. It is to be noted that it is not sufficient to define the immunogen as being derivable from a method claimed (see PCT Guidelines, Part II, Chapter 5, Appendix A5.26 and A5.26[1]). This remark also applies to claims 34-38 when relating to claim 33.

Industrial applicability (Art. 33(4) PCT)

7. For the assessment of the present claims 36-38 on the question whether they are industrially applicable, no unified criteria exist in the PCT Contracting States. The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment. The remaining claims 1-35 meet the requirements of industrial applicability.

Further comments

8. The above comments are based on the assumption that all claims enjoy priority rights from the filing date of the priority document. In that case the document "WO 02/070711" cited as P,X-document in the international search report is not considered as prior art under Article 33 PCT.
9. For the above cited prior art documents and the references in the description, see Rule 5.1(a)(ii) PCT).

REPLACED BY
ART 34 AND
Pages 1-48 of
original Spec.

Vaccine

The present invention relates to isolated immunogens and their use in the treatment of diseases that are treatable with neutralisation of IL-13, such as COPD, asthma and atopic disorders such as hayfever, contact allergies and atopic dermatitis. Most preferably the immunogens are used in the treatment of asthma. In particular the invention relates to the neutralisation of the biological effects of IL-13 by raising an immune response against the IL-13 by vaccination of a mammal with immunogens comprising the native or mutated amino acid sequence of IL-13, and foreign T-helper epitopes either inserted in, or attached to the IL-13 sequence or present in carrier polypeptides. Also provided by the present invention are DNA vaccines that comprise a polynucleotide sequence that encodes the immunogens of the present invention. The invention further relates to pharmaceutical compositions comprising such immunogens and their use in medicine and to methods for their production.

Background to the invention

COPD is an umbrella term to describe diseases of the respiratory tract, which shows similar symptoms to asthma and is treated with the same drugs. COPD is characterised by a chronic, progressive and largely irreversible airflow obstruction. The contribution of the individual to the course of the disease is unknown, but smoking cigarettes is thought to cause 90% of the cases. Symptoms include coughing, chronic bronchitis, breathlessness and respiratory infections. Ultimately the disease will lead to severe disability and death.

Asthma is a chronic lung disease, caused by inflammation of the lower airways and is characterised by recurrent breathing problems. Airways of patients are sensitive and swollen or inflamed to some degree all the time, even when there are no symptoms. Inflammation results in narrowing of the airways and reduces the flow of air in and out of the lungs, making breathing difficult and leading to wheezing, chest tightness and coughing. Asthma is triggered by super-sensitivity towards allergens (e.g. dust mites, pollens, moulds), irritants (e.g. smoke, fumes, strong odours), respiratory infections, exercise and dry weather. The triggers irritate the airways and the lining of the airways swell to become even more inflamed, mucus then clogs up the airways and the muscles around the airways tighten up until breathing becomes difficult and stressful and asthma symptoms appear.

Atopic disorders refers to a group of diseases that are hereditary and often occur together, including asthma, allergies such as hay fever, and atopic dermatitis. Atopic

dermatitis is a chronic disease that affects the skin. In atopic dermatitis, the skin becomes extremely itchy and inflamed, causing redness, swelling, cracking, weeping, crusting, and scaling. Atopic dermatitis most often affects infants and young children, but it can continue into adulthood or first show up later in life. In most cases, there are periods of time when the disease is worse, called exacerbations or flares, followed by periods when the skin improves or clears up entirely, called remissions. Many children with atopic dermatitis will experience a permanent remission of the disease when they get older, although their skin often remains dry and easily irritated. Environmental factors can bring on symptoms of atopic dermatitis at any time in the lives of individuals who have inherited the atopic disease trait. Atopic dermatitis is often referred to as "eczema," which is a general term for the many types of dermatitis. Atopic dermatitis is the most common of the many types of eczema. Several have very similar symptoms.

The way the skin is affected by atopic dermatitis can be changed by patterns of scratching and resulting skin infections. Some people with the disease develop red, scaling skin where the immune system in the skin is becoming very activated. Others develop thick and leathery skin as a result of constant scratching and rubbing. This condition is called lichenification. Still others develop papules, or small raised bumps, on their skin. When the papules are scratched, they may open (excoriations) and become crusty and infected.

Many factors or conditions can make symptoms of atopic dermatitis worse, further triggering the already overactive immune system in the skin, aggravating the itch-scratch cycle, and increasing damage to the skin. These exacerbating factors can be broken down into two main categories: irritants (such as wool or synthetic fibers, rough or poorly fitting clothing, soaps and detergents, some perfumes and cosmetics, chlorine, mineral oil, some solvents, dust or sand) and allergens (such as pollen, dog or cat dander, and dust mite allergens). Emotional factors and some infections can also influence atopic dermatitis.

If a flare of atopic dermatitis does occur, several methods can be used to treat the symptoms. Corticosteroids as topical creams are the most frequently used treatment, although systemic administration is also used in some severe cases. Sometimes over-the-counter preparations are used, but in many cases the doctor will prescribe a stronger corticosteroid cream or ointment. An example of a commonly prescribed corticosteroid is prednisone. Side effects of repeated or long-term use of topical corticosteroids can include thinning of the

skin, infections, growth suppression (in children), and stretch marks on the skin. Antibiotics to treat skin infections may be applied directly to the skin in an ointment, but are usually more effective when taken by mouth. Phototherapy (treatment with light) that uses ultraviolet A or B light waves, or both together, can be an effective treatment for mild to moderate
5 dermatitis in older children (over 12 years old) and adults. In adults, immunosuppressive drugs, such as cyclosporine, are also used to treat severe cases of atopic dermatitis that have failed to respond to any other forms of therapy. The side effects of cyclosporine can include high blood pressure, nausea, vomiting, kidney problems, headaches, tingling or numbness, and a possible increased risk of cancer and infections.

10 Because of the unmet medical need therefor and the side affects of existing therapies there is a need for alternative treatments for atopic diseases in general, and in particular for treatments for asthma and atopic dermatitis.

IL-13 is a Th2-type cytokine that is closely related to IL-4. A number of recent papers have defined the role for IL-13 in driving pathology in the ovalbumin model of allergenic
15 asthma (Wills-Karp et al, 1998, *Science* 282:2258-2261; Grunig et al, 1998, *Science* 282:2261-2263). In this work, mice previously sensitised to ovalbumin were injected with a soluble IL-13 receptor which binds and neutralises IL-13. Airway hyper-responsiveness to acetylcholine challenge was reduced in the treated group. Histological analysis revealed that treated mice had reversed the goblet-cell metaplasia seen in controls. In complementary
20 experiments, lung IL-13 levels were raised by over-expression in a transgenic mouse or by installation of protein into the trachea in wild-type mice. In both settings, airway hyper-responsiveness, eosinophil invasion and increased mucus production were seen (Zhu et al, 1999, *J.Clin.Invest.* 103:779-788).

The sequence of the mature form of human IL-13 is provided in SEQ ID No. 1 and is
25 shown in FIG. 1.

The sequence of the mature form of murine IL-13 is provided in SEQ ID No. 2 and is shown in FIG. 2.

Sequences for IL-13 from several mammalian species and non-human primates are shown in FIG. 3 and FIG. 4 (SEQ ID NO.s 3 to 9)

30 As a result of the various problems associated with the production, administration and tolerance of monoclonal antibodies there is an increased focus on methods of instructing the

patient's own immune system to generate endogenous antibodies of the appropriate specificity by means of vaccination. However, mammals do not generally have high-titre antibodies against self-proteins present in serum, as the immune system contains homeostatic mechanisms to prevent their formation. The importance of these "tolerance" mechanisms is
5 illustrated by diseases like myasthenia gravis, in which auto-antibodies directed to the nicotinic acetylcholine receptor of skeletal muscle cause weakness and fatigue (Drachman, 1994, *N Engl J Med* 330:1797-1810).

A number of techniques have been designed with the aim of breaking "tolerance" to self antigen. One technique involves chemically cross-linking the self-protein (or peptides
10 derived from it) to a highly immunogenic carrier protein, such as keyhole limpet haemocyanin ("Antibodies: A laboratory manual" Harlow, E and Lane D. 1988. Cold Spring Harbor Press).

A variant on the carrier protein technique involves the construction of a gene encoding a fusion protein comprising both carrier protein (for example hepatitis B core
15 protein) and self-protein (The core antigen of hepatitis B virus as a carrier for immunogenic peptides", *Biological Chemistry*. 380(3):277-83, 1999). The fusion gene may be administered directly as part of a nucleic acid vaccine. Alternatively, it may be expressed in a suitable host cell *in vitro*, the gene product purified and then delivered as a conventional vaccine, with or without an adjuvant.

20 Another approach has been described by Dalum and colleagues wherein a single class II MHC-restricted epitope is inserted into the target molecule. They demonstrated the use of this method to induce antibodies to ubiquitin (Dalum et al, 1996, *J Immunol* 157:4796-4804; Dalum et al, 1997, *Mol Immunol* 34:1113-1120) and the cytokine TNF (Dalum et al, 1999, *Nature Biotech* 17:666-669). As a result, all T cell help must arise either from this single
25 epitope or from junctional sequences. Such an approach is also described in EP 0 752 886 B1, WO 95/05849, and WO 00/65058.

Treatment therapies, some including vaccination, for the neutralisation of several cytokines are known. WO 00/65058 describes a method of down regulating the function of the cytokine IL-5, and its use in the treatment of asthma. In this study, the IL-5 sequence was
30 modified by a number of techniques to render it immunogenic, amongst which there is described an IL-5 immunogen supplemented with foreign T-cell epitopes, whilst maintaining

the IL-5 B cell epitopes. WO 01/62287 discloses IL-13, amongst a long list of potential antigens, for use in allergy or asthma vaccines. WO 00/06937 discloses cytokine derivatives that are functionally inactivated for use as vaccine antigens. Chimaeric IL-13 immunogens are disclosed in the co-pending patent application WO 02/070711.

5 There remains a need to provide improved immunotherapeutic treatments for asthma, and improved immunogens for raising neutralising anti-IL-13 immune responses.

Summary of the Invention

The present invention provides pharmaceutical compositions comprising modified “self” IL-13 immunogens, wherein the IL-13 immunogen is modified to include foreign T-
10 cell helper epitopes. The pharmaceutical composition is preferably for use in human therapy, and in this composition the IL-13 sequence is a human sequence or other sequence that is capable of generating an immune response that recognises human IL-13; and the T-cell helper epitopes are “foreign” with respect to human self-proteins. Preferably the T-helper epitopes are also foreign with respect to other IL-13 sequences from other species. However,
15 animal pharmaceutical products are not excluded, for example canine or other veterinary species pharmaceutical products can be made in an analogous fashion to that described for human vaccines above.

The compositions of the present invention comprise an IL-13 element and an additional element for providing T-cell help.

20 *IL-13 element*

The IL-13 element, in its broadest form, is any sequence that is capable of driving an immune response that recognises and neutralises the biological effects of IL-13. Preferably, the IL-13 is human IL-13.

In this context of the present invention the entire IL-13 sequences may be used, or
25 functional equivalent fragments thereof. Accordingly, references in this text to IL-13 sequences may encompass the entire sequence or fragments or truncates thereof.

The IL-13 element may comprise the native IL-13 sequence or a mutated form thereof. Accordingly, the IL-13 sequence may be, for example, native human IL-13 or fragment thereof.

30 In an alternative embodiment of the present invention the immunogens comprise a chimaeric IL-13 sequence that comprise substitution mutations to swap one or more of the

human sequence amino acids with the equivalent amino acids found in the same positions within the sequence of IL-13 from another mammalian species. In the context of a human vaccine immunogen, the object of the chimaeric sequences is to maximise the amino acid sequence diversity between the immunogen and human native IL-13, whilst keeping maximal shape and conformational homology between the two compositions. The chimaeric immunogen achieves this by substituting amino acids found in regions predicted to be masked from the surface. Most preferably the amino acids are substituted with amino acids that are found in equivalent positions within an IL-13 sequence from another mammalian species. In this way, sequence diversity is achieved with minimal alteration to the overall shape/configuration of the immunogen.

In one aspect of the present invention, there is provided a human IL-13 immunogen that comprises substitution mutations in areas that are associated with alpha helical regions, which substitutions involve swapping the human amino acid with the amino acid that appears in the same position within the IL-13 sequence of a different mammalian species.

Most preferably, there are substitution mutations in a plurality of sites within the IL-13 sequence, wherein at least two or more of the mutation sites comprise a substitution involving amino acids taken from different non-human mammalian species, more preferably the substitutions involve amino acids taken from 3 or more different non-human mammalian species, and most preferably the substitutions involve amino acids taken from 4 or more different non-human mammalian species.

Preferably, the substitutions do not occur in at least six of the areas of high interspecies conservation: 3PVP, 12ELIEEL, 19NITQ, 28LCN, 32SMVWS, 50SL, 60AI, 64TQ, 87DTKIEVA, 99LL, 106LF.

The preferred IL-13 element of the immunogens of the present invention are human chimaeric IL-13 sequences which have a similar conformational shape to native human IL-13 whilst having sufficient amino acid sequence diversity to enhance its immunogenicity when administered to a human, characterised in that the chimaeric IL-13 immunogen has the sequence of human IL-13 comprising:

(a) substitution mutations in at least two of the following alpha helical regions:

PSTALRELIEELVNIT, MYCAALESIL, KTQRMLSGF or AQFVKDLLLHLKKLFRF,

(b) comprises in unmutated form at least six of the following regions of high inter-species conservation 3PVP, 12ELIEEL, 19NITQ, 28LCN, 32SMVWS, 50SL, 60AI, 64TQ, 87DTKIEVA, 99LL, 106LF, and

(c) optionally comprises a mutation in any of the remaining amino acids,

5 wherein any substitution performed in steps a, b or c is a structurally conservative substitution.

The numerical prefix to the amino acids listed, refers to the positional number of the amino acid sequence in the mature form of human IL-13, wherein the first residue "G" is assigned the number 2.

10 In the context of step (a) of the above chimaeric IL-13 element, preferably at least two, more preferably at least three and most preferably all four alpha helical regions comprise at least one substitution mutation. In the context of step (b) preferably at least 7, more preferably at least 8, more preferably at least 9, more preferably at least 10, and most preferably all 11 of the regions are unmutated.

15 Preferably greater than 50% of these substitutions or mutations in the above chimaeric IL-13 element, comprise amino acids taken from equivalent positions within the IL-13 sequence of a non-human. More preferably more than 60, or 70, or 80 percent of the substitutions comprise amino acids taken from equivalent positions within the IL-13 sequence of a non-human mammal. Most preferably, each substitution or mutation comprise
20 amino acids taken from equivalent positions within the IL-13 sequence of a non-human mammal.

Again in the context of the chimaeric human IL-13 element, preferably greater than 50% of these substitutions or mutations occur in regions of human IL-13 which are predicted to be alpha helical in configuration. More preferably more than 60, or 70, or 80 percent of the
25 substitutions or mutations occur in regions of human IL-13 which are predicted to be alpha helical in configuration. Most preferably, each substitution or mutation occurs in regions of human IL-13 which are predicted to be alpha helical in configuration.

Again in the context of the chimaeric human IL-13 elements, preferably the human IL-13 sequence comprises between 2 and 20 substitutions, more preferably between 6 and 15
30 substitutions and most preferably 13 substitutions.

In the case of a human IL-13 vaccine, the IL-13 immunogen could be based on an orthologous IL-13 sequence (such as the murine IL-13 sequence) wherein the murine B-cell epitopes (surface exposed regions) are substituted for the equivalent human sequences. In this embodiment the murine "backbone" will provide foreign T-cell epitopes, in addition to the supplemental promiscuous T-cell epitopes (such as P2 or P30) which are added either at the termini or within the chimaera sequence.

A preferred chimaeric human IL-13 immunogen comprises the sequence of human IL-13, wherein the amino acid sequence comprises conservative substitutions, or substitutions characteristic of amino acids present at equivalent positions within the IL-13 sequence of a non-human species, present in at least six of the following 13 positions 8T, 11R, 18V, 49E, 62K, 66M, 69G, 84H, 97K, 101L, 105K, 109E, 111R. Most preferably such a chimaeric human IL-13 immunogen comprises at least 6, and preferably all, of the following substitutions:

Position	Substitution	Species
8	T->S	Synthetic
11	R->K	pig, cow, dog, mouse, gerbil, cyno, rhesus, marmoset.
18	V->A	Synthetic
49	E->D	cow, mouse, gerbil.
62	K->R	cow, dog, mouse, rat.
66	M->I	Mouse, gerbil, rat.
69	G->A	Cow, pig, dog
84	H->R	Dog, rhesus, cyno
97	K->T	Mouse
101	L->V	Cyno, rhesus
105	K->R	Synthetic
109	E->Q	Marmoset
111	R->T	Marmoset

The chimaeric IL-13 that comprises each of these listed substitutions is a preferred IL-13 element (Immunogen 1, SEQ ID NO. 10) and is shown in FIG. 5. Other highly

preferred IL-13 elements are Immunogen 11 (SEQ ID NO. 20, see FIG 15), Immunogen 12 (SEQ ID NO. 21, see FIG. 16) and Immunogen 13 (SEQ ID NO. 22, see FIG. 17).

The IL-13 element may also optionally further comprise a mutation that abolishes the biological activity of the immunogen. The following substitutions can be used to inactivate human IL13 bioactivity: E 12 to I, S, or Y; E12 to K; R 65 to D; S 68 to D; R 108 to D.

In certain aspects of the present invention immunogenic fragments of the native IL-13 sequence may be used, for example in the presentation of immunogenic peptides in Hepatitis B core particles or in the context of chimaeric immunogens described above. In these contexts immunogenic fragments of the human IL-13 sequences preferably contain the B-cell epitopes in the human IL-13 sequence, and preferably at least one or more of the following short sequences:

GPVPPSTA
ITQNQKAPLCNGSMVWSINLTAGM
INSGCS
15 FCPHKVSAGQFSSLHVRDT
LHLKKLFREGRFN

The polypeptide of the invention may be further modified by mutation, for example substitution, insertion or deletion of amino-acids in order to add desirable properties (such as the addition of a sequence tag that facilitates purification or increase immunogenicity) or remove undesirable properties (such as an unwanted agonistic activity at a receptor) or trans-membrane domains. In particular the present invention specifically contemplates fusion partners that ease purification such as poly histidine tags or GST expression partners that enhance expression. A preferred tag or expression partner is immunoglobulin FC of human IgG1 fused to the C-terminus of the IL-13 molecule.

Other mutations, outside of those regions that are to be left unmutated due to their high level of conservation between species, may occur in the IL-13 sequence. Preferably such mutations are conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged.

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such

as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless
5 obtain a protein with like properties. It is thus contemplated that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

In making such changes, the hydropathic index of amino acids may be considered.
10 The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates,
15 receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6);
20 histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, i.e. still obtain a biological functionally equivalent protein. In making such changes,
25 the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average
30 hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (−0.4); proline (−0.5 \pm 1); alanine (−0.5); histidine (−0.5); cysteine (−1.0);
5 methionine (−1.3); valine (−1.5); leucine (−1.8); isoleucine (−1.8); tyrosine (−2.3); phenylalanine (−2.5); tryptophan (−3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those
10 within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take
15 various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine. These are preferred conservative substitutions.

Amino acid substitutions may further be made on the basis of similarity in polarity,
20 charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, &
25 phenylalanine and tyrosine. Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his.

Element to provide T-cell help.

Associated with the IL-13 element to make the immunogens of the present invention, are elements that provide foreign T-cell help. Most preferably the T-cell helper epitopes are foreign to human sequences, but also foreign with respect to any IL-13 sequences from non-human mammals. Preferably the T-cell helper epitopes used are small and are added to the
5 IL-13 sequence by an addition or substitution event within, or at the terminal ends of, the IL-13 sequence by synthetic, recombinant or molecular biological means. Alternatively the T-cell helper epitopes may be added via chemical coupling of the IL-13 polypeptide to a carrier protein comprising the T-cell helper epitopes. The IL-13 sequences, or functionally equivalent fragments thereof, may also be associated with the T-cell helper epitopes in a
10 fusion protein, wherein the two are recombinantly manufactured together, for example a Hepatitis B core protein incorporating IL-13 sequences.

In the aspects of the present invention where small T-cell helper epitopes are used, a "foreign T-cell helper epitope" or "T-cell epitope" is a peptide which is able to bind to an MHC II molecule and stimulates T-cells in an animal species. Preferred foreign T-cell
15 epitopes are promiscuous epitopes, *ie.* epitopes that bind multiple different MHC class II molecules in an animal species or population (Panina-Bordignon et al, *Eur.J.Immunol.* 1989, 19:2237-2242; Reece et al, *J.Immunol.* 1993, 151:6175-6184; WO 95/07707).

In order for the immunogens of the present invention to be clinically effective in a complex outbred human population, it may be advantageous to include several foreign T-cell
20 epitopes. Promiscuous epitopes may also be another way of achieving this same effect, including naturally occurring human T-cell epitopes such as those from tetanus toxoid (e.g. the P2 and P30 epitopes, diphtheria toxoid, influenza virus haemagglutinin (HA), and *P.falciparum* CS antigen. The most preferred T-cell epitopes for use in the present invention are P2 and P30 from tetanus toxoid

25 A number of promiscuous T-cell epitopes have been described in the literature, including: WO 98/23635; Southwood et al., 1998, *J. Immunol.*, 160: 3363-3373; Sinigaglia et al., 1988, *Nature*, 336: 778-780; Rammensee et al., 1995, *Immunogenetics*, 41: 4, 178-228; Chicz et al., 1993, *J. Exp. Med.*, 178:27-47; Hammer et al., 1993, *Cell* 74:197-203; and Falk et al., 1994, *Immunogenetics*, 39: 230-242. The promiscuous T-cell epitope can also be an
30 artificial sequence such as "PADRE" (WO 95/07707).

The heterologous T-cell epitope is preferably selected from the group of epitopes that will bind to a number of individuals expressing more than one MHC II molecules in humans. For example, epitopes that are specifically contemplated are P2 and P30 epitopes from tetanus toxoid, Panina – Bordignon Eur. J. Immunol 19 (12), 2237 (1989). In a preferred
 5 embodiment the heterologous T-cell epitope is P2 or P30 from Tetanus toxin.

The P2 epitope has the sequence QYIKANSKFIGITE and corresponds to amino acids 830-843 of the Tetanus toxin.

The P30 epitope (residues 947-967 of Tetanus Toxin) has the sequence FNNFTVSFWLRVPKVSASHLE. The FNNFTV sequence may optionally be deleted.
 10 Other universal T epitopes can be derived from the circumsporozoite protein from Plasmodium falciparum – in particular the region 378-398 having the sequence DIEKKIAKMEKASSVFNVVNS (Alexander J, (1994) Immunity 1 (9), p 751-761). Another epitope is derived from Measles virus fusion protein at residue 288-302 having the sequence LSEIKGVIVHRLEGV (Partidos CD, 1990, J. Gen. Virol 71(9) 2099-2105).
 15 Yet another epitope is derived from hepatitis B virus surface antigen, in particular amino acids, having the sequence FLLTRILTIPQSLD.

Another set of epitopes is derived from diptheria toxin. Four of these peptides (amino acids 271-290, 321-340, 331-350, 351-370) map within the T domain of fragment B of the toxin, and the remaining 2 map in the R domain (411-430, 431-450):

20 PVFAGANYAAWAVNVAQVI
 VHHNTEEIVAQSIALSSLMV
 QSIALSSLMVAQAIPLVGEL
 VDIGFAAYNFVESII NLFQV
 QGESGHDIKITAENTPLPIA
 25 GVLLPTIPGKLDVNKSKTHI

(Raju R., Navaneetham D., Okita D., Diethelm-Okita B., McCormick D., Conti-Fine B. M. (1995) Eur. J. Immunol. 25: 3207-14.)

A particularly preferred element to provide T-cell help, is a fusion partner called "CPC" (clyta-P2-clyta) which is disclosed in PCT/EP03/06096.

Most preferably the foreign T-cell helper epitopes are "foreign" in that they are not tolerated by the host immune system, and also in that they are not sequences that are derived or selected from any IL-13 sequence from another species (non-vaccinee).

In the aspect of the present invention where native self IL-13 is coupled to a T-helper epitope bearing immunogenic carrier, the conjugation can be carried out in a manner well known in the art. Thus, for example, for direct covalent coupling it is possible to utilise a carbodiimide, glutaraldehyde or (N-[γ -maleimidobutyryloxy] succinimide ester, utilising common commercially available heterobifunctional linkers such as CDAP and SPDP (using manufacturers instructions). After the coupling reaction, the immunogen can easily be isolated and purified by means of a dialysis method, a gel filtration method, a fractionation method etc.

The types of carriers used in the immunogens of the present invention will be readily known to the man skilled in the art. A non-exhaustive list of carriers which may be used in the present invention include: Keyhole limpet Haemocyanin (KLH), serum albumins such as bovine serum albumin (BSA), inactivated bacterial toxins such as tetanus or diphtheria toxins (TT and DT), or recombinant fragments thereof (for example, Domain 1 of Fragment C of TT, or the translocation domain of DT), or the purified protein derivative of tuberculin (PPD). Alternatively the IL-13 may be directly conjugated to liposome carriers, which may additionally comprise immunogens capable of providing T-cell help. Preferably the ratio of IL-13 to carrier molecules is in the order of 1:1 to 20:1, and preferably each carrier should carry between 3-15 IL-13 molecules.

In an embodiment of the invention a preferred carrier is Protein D from *Haemophilus influenzae* (EP 0 594 610 B1). Protein D is an IgD-binding protein from *Haemophilus influenzae* and has been patented by Forsgren (WO 91/18926, granted EP 0 594 610 B1). In some circumstances, for example in recombinant immunogen expression systems it may be desirable to use fragments of protein D, for example Protein D 1/3rd (comprising the N-terminal 100-110 amino acids of protein D (GB 9717953.5)).

Another preferred method of presenting the IL-13, or immunogenic fragments thereof, is in the context of a recombinant fusion molecule. For example, EP 0 421 635 B describes the use of chimaeric hepadnavirus core antigen particles to present foreign peptide sequences in a virus-like particle. As such, immunogens of the present invention may

comprise IL-13 presented in chimaeric particles consisting of hepatitis B core antigen. Additionally, the recombinant fusion proteins may comprise IL-13 and a carrier protein, such as NS1 of the influenza virus. For any recombinantly expressed protein which forms part of the present invention, the nucleic acid which encodes said immunogen also forms an aspect
5 of the present invention.

Preferred Immunogens

In the sections above, preferred definitions of the IL-13 element and the element to provide T-cell help have been described. For the compositions of the present invention, it is intended that this document discloses each individual preferred element from the IL-13
10 element section in combination with each individual preferred element from the element to provide T-cell help section. Particularly preferred are combinations of Immunogens 1, 11, 12 or 13, and a carrier protein or promiscuous T-cell helper epitope. Preferred carrier protein or promiscuous T-cell helper epitopes include Protein D, CPC, P2 or P30.

Specifically disclosed preferred combinations of elements to form preferred
15 immunogens are listed herebelow.

When the IL-13 element is native human IL-13, and the element that provides T-cell help is a promiscuous T-cell epitope, preferred examples include: Immunogen 2 (see FIG. 6, SEQ ID NO. 11), which comprises human IL-13 with P30 inserted (underlined) into the protein (substituted for the looped region between alpha helices C and D of human IL13).

20 Immunogen 3 (FIG. 7, SEQ ID NO. 12) is a Human IL-13 immunogen with N-terminal P30.

Immunogen 4 (FIG. 8, SEQ ID NO. 13) is a murine IL-13 with p30 inserted into the protein (substituted for the looped region between alpha helices C and D of mouse IL13) this is an example of a mouse version of an IL13 autovaccine. The p30 region is underlined.

25 Immunogen 5 (FIG. 9, SEQ ID NO. 14) is a murine IL13 with p30 at the N-terminus. This is an example of a mouse version of an IL13 autovaccine. The p30 region is underlined and is positioned at the N-terminus of the mature mouse IL13 protein sequence.

Specific examples where the IL-13 element is provided as a chimaeric IL-13 immunogen include:

Immunogen 6 (FIG. 10, SEQ ID NO. 15). This is an example of a mouse version of this form of the vaccine, where there is "human backbone" sequence grafted to murine B-cell surface exposed epitopes, with P30 added at the N-terminus.

Other preferred immunogens are based on a human chimaeric IL-13 "Immunogen 1" (SEQ ID NO. 10). For example, Immunogen 1 is preferably N-terminally fused to the carrier "CPC" to form Immunogen 7 (SEQ ID NO. 16, see FIG. 11), or N-terminally fused to protein D (the protein D fusion region corresponds to amino acids S20 to T127 inclusive, of H.influenzae protein D sequence (nb, the DNA sequence encoding the protein D is codon optimised) for Immunogen 8 (SEQ ID NO. 17, see FIG. 12); or N-terminally fused to P30 to give Immunogen 9 (SEQ ID NO.18, see FIG. 13). Immunogen 9 preferably further comprises the E121 mutation to abrogate any IL-13 biological activity, to give Immunogen 10 (SEQ ID NO. 19, see FIG. 14).

The protein and DNA sequences shown for Immunogens 1 to 10 are shown without the amino acid or DNA sequence for the signal sequence required to drive secretion of the product from the cell. Preferably, therefore, the sequences further are further provided with a signal sequence. In the context of DNA vaccines it is specifically preferred that the signal sequence is a non-human derived sequence that comprises a T-cell epitope, to further provide T-cell help. None of the disclosed preferred sequences have a stop codon as it may be useful to express them fused to other molecules eg immunoglobulin Fc, 6His to facilitate production or purification.

The numbering system used herein conforms with normal practice in the field of IL-13, in that the G in "GPVPP" is referred to as residue 2, and the remaining amino acids are numbered accordingly.

25 *Methods of designing a vaccine*

In an important aspect of the present invention, there is provided a method of designing a vaccine for the treatment of an individual suffering from or susceptible to a disease that is susceptible to treatment by neutralisation of the activity of IL-13. Such diseases include COPD, asthma and atopic disorders such as hayfever, contact allergies and atopic dermatitis.

The methods disclosed herein comprise two major steps: 1. Designing a chimaeric IL-13 immunogen, and 2. Associating to the IL-13 immunogen, a source of T-cell epitopes that are foreign with respect to any human self epitope and also foreign with respect to any mammalian IL-13 sequence.

5 In this context the method comprises:

(a) taking the sequence of human IL-13 and identifying regions that are predicted to form an alpha helical structure, and

(b) mutating the sequence of human IL-13 within these alpha helical regions to substitute amino acids from the human sequence with amino acids that are either a
10 conservative substitution or are found in equivalent positions within the IL-13 sequence of a different species, and

c) attaching or inserting a source of T-cell epitopes that are foreign with respect to any human self epitope and also foreign with respect to any mammalian IL-13 sequence.

As a general principle the object of the method is to design a chimaeric sequence
15 having a maximum sequence diversity between the immunogen and human native IL-13, whilst keeping maximal shape and conformational homology between the two compositions. The chimaeric immunogen achieves this by substituting amino acids found in regions predicted to be masked from the surface. Most preferably the amino acids are substituted with amino acids that are found in equivalent positions within an IL-13 sequence from
20 another mammalian species. In this way, sequence diversity is achieved with minimal alteration to the overall shape/configuration of the immunogen.

Therefore, the preferred methods of designing a chimaeric IL-13 immunogen comprise the following steps:

1. Collect together IL13 sequences from other and align using tool such as Clustal or Pileup,
- 25 2. Avoid mutations within positions which are essentially invariant across the collection. Particularly 3PVP, 12ELIEEL, 19NITQ, 28LCN, 32SMVWS, 50SL, 60AI, 64TQ, 87DTKIEVA, 99LL, 106LF,
3. In the remaining sequence, favour mutations that occur in the helical regions (PSTALRELIEELVNIT, MYCAALESIL, KTQRMLSGF, AQFVKDLLLHLKCLFRE)
- 30 4. Regions not specified in 3 or 4 may optionally contain mutations.

5. Mutations are selected by considering either residues which occur in other species IL13 molecules at orthologous positions, or those which are chemically conservative.

Molecular modelling may be used to select particularly favourable substitutions which have a low probability of affecting the overall shape of the molecule by steric clashes
5 etc.

Accordingly there is provided a method for the manufacture of a human chimaeric IL-13 immunogen which has a similar conformational shape to native human IL-13 whilst having sufficient amino acid sequence diversity to enhance its immunogenicity when administered to a human, the method comprising the following steps:

- 10 (a) taking the sequence of human IL-13 and performing at least one substitution mutation in at least two of the following alpha helical regions: PSTALRELIEELVNIT, MYCAALESLLI, KTQRMLSGF or AQFVKDLLLHLKKLFRE,
(b) preserving at least six of the following regions of high inter-species conservation 3PVP, 12ELIEEL, 19NITQ, 28LCN, 32SMVWS, 50SL, 60AI, 64TQ, 87DTKIEVA, 99LL, 106LF,
15 (c) optionally mutating any of the remaining amino acids, and
(d) attaching a source of T-cell epitopes that are foreign with respect to any human self epitope and also foreign with respect to any mammalian IL-13 sequence, characterised in that any substitution performed in steps a, b or c is a structurally conservative substitution.

- 20 In the context of step (a) preferably at least two, more preferably at least three and most preferably all four alpha helical regions comprise at least one substitution mutation. In the context of step (b) preferably at least 7, more preferably at least 8, more preferably at least 9, more preferably at least 10, and most preferably all 11 of the regions are unmutated.

Alternatively there is provided, a method for the manufacture of a human chimaeric
25 IL-13 immunogen which has a similar conformational shape to native human IL-13 whilst having sufficient amino acid sequence diversity to enhance its immunogenicity when administered to a human, the method comprising the following steps:

- (a) aligning IL-13 amino acid sequences from different species,
(b) identifying regions of high variability and high conservation,

- (c) taking the sequence of human IL-13 and mutating it in the areas of high variability to substitute amino acids from the human sequence with amino acids that are either a conservative substitution or are found in equivalent positions within the IL-13 sequence of a different species, and
- 5 (d) attaching a source of T-cell epitopes that are foreign with respect to any human self epitope and also foreign with respect to any mammalian IL-13 sequence,

In a related aspect of the present invention, there is also provided a method for the manufacture of a human chimaeric IL-13 immunogen comprising the following steps:

- 10 (a) aligning IL-13 amino acid sequences from different species,
(b) identifying regions of high variability and high conservation,
(c) taking the sequence of human IL-13 and mutating it in the areas of high conservation to substitute amino acids from the human sequence with amino acids that are either a conservative substitution or are found in equivalent positions within the IL-13 sequence of a
15 different species, and
(d) attaching a source of T-cell epitopes that are foreign with respect to any human self epitope and also foreign with respect to any mammalian IL-13 sequence,

In all of these methods, preferably greater than 50% of these substitutions or
20 mutations comprise amino acids taken from equivalent positions within the IL-13 sequence of a non-human. More preferably more than 60, or 70, or 80 percent of the substitutions comprise amino acids taken from equivalent positions within the IL-13 sequence of a non-human mammal. Most preferably, each substitution or mutation comprise amino acids taken from equivalent positions within the IL-13 sequence of a non-human mammal.

- 25 Again in the context of the methods for designing chimaeric human immunogens, preferably greater than 50% of these substitutions or mutations occur in regions of human IL-13 which are predicted to be alpha helical in configuration. More preferably more than 60, or 70, or 80 percent of the substitutions or mutations occur in regions of human IL-13 which are predicted to be alpha helical in configuration. Most preferably, each substitution or mutation
30 occurs in regions of human IL-13 which are predicted to be alpha helical in configuration.

Again in the context of the methods of designing chimaeric human immunogens, preferably the immunogen comprises between 2 and 20 substitutions, more preferably between 6 and 15 substitutions, and most preferably 13 substitutions.

Most preferably, in all of these above methods there are substitution mutations in a plurality of sites within the IL-13 sequence, wherein at least two or more of the mutation sites comprise a substitution involving amino acids taken from different non-human mammalian species, more preferably the substitutions involve amino acids taken from 3 or more different non-human mammalian species, and most preferably the substitutions involve amino acids taken from 4 or more different non-human mammalian species.

10 The present invention also provides an immunogen that is derivable from any of the above methods, which immunogens are immunogenic, when formulated in an appropriate manner for a vaccine, in a human vaccinee.

The successful design of a polypeptide according to the present invention can be verified for example by administering the resulting polypeptide in a self-context in an appropriate vaccination regime, and observing that antibodies capable of binding the protein are induced. This binding may be assessed through use of ELISA techniques employing recombinant or purified native protein, or through bioassays examining the effect of the protein on a sensitive cell or tissue. A particularly favoured assessment is to observe a phenomenon causally related to activity of the protein in the intact host, and to determine whether the presence of antibodies induced by the methods of the invention modulate that phenomenon. Thus a protein of the present invention will be able to raise antibodies to the native antigen in the species from which the native protein is derived.

20 The most successful of designs will be able to be used in an experiment, such as that described in Example 2 herein, and induce anti-IL-13 neutralising immune responses that exceed ED100 in at least 50% of the vaccinated individuals.

Vaccine formulations

The vaccine formulations of the present invention may be in the form of a protein based vaccine, most often formulated together with an adjuvant, or alternatively the vaccine may take the form of a DNA or polynucleotide vaccine.

The polypeptide immunogens of the invention may be encoded by polynucleotides of the invention. A person skilled in the art will readily be able to determine the sequence of the polynucleotide which encodes the polypeptide by applying the genetic code. Once the required nucleic acid sequence has been determined, the polynucleotide with the desired
5 sequence can be produced as described in the examples. A skilled person will readily be able to adapt any parameters necessary, such as primers and PCR conditions. It will also be understood by a person skilled in the art that, due to the degeneracy of the genetic code, there is potentially more than one polynucleotide which encodes a polypeptide of the invention. The polynucleotides of the present invention may also comprise a region which encodes a
10 secretion signal peptide.

The polynucleotide of the invention is typically RNA, for example mRNA, or DNA, for example genomic DNA, cDNA or synthetic DNA. Preferably the polynucleotide is DNA. Particularly preferably it is cDNA.

The present invention further provides an expression vector, which is a nucleic acid
15 construct, comprising the polynucleotide of the invention. Additionally, the nucleic acid construct will comprise appropriate initiators, promoters, enhancers and other elements, such as for example, polyadenylation signals, which may be necessary, and which are positioned in the correct orientation, in order to allow for protein expression within a mammalian cell.

The promoter may be a eukaryotic promoter for example a CD68 promoter, Gal1,
20 Gal10, or NMT1 promoter, a prokaryotic promoter for example Tac, Trc, or Lac, or a viral promoter, for example the cytomegalovirus promoter, the SV40 promoter, the polyhedrin promoter, the P10 promoter, or the respiratory syncytial virus LTR promoter. Preferably the promoter is a viral promoter. Particularly preferred is when the promoter is the cytomegalovirus immediate early promoter, optionally comprising exon 1 from the HCMV
25 IE gene.

The transcriptional regulatory elements may comprise enhancers, for example the hepatitis B surface antigen 3'untranslated region, the CMV enhancer; introns, for example the CD68 intron, or the CMV intron A, or regulatory regions, for example the CMV 5' untranslated region.

The polynucleotide is preferably operably linked to the promoter on the nucleic acid construct such that when the construct is inserted into a mammalian cell, the polynucleotide is expressed to produce a encoded polypeptide.

The nucleic acid construct backbone may be RNA or DNA, for example plasmid DNA, viral DNA, bacterial DNA, bacterial artificial chromosome DNA, yeast artificial chromosome DNA, synthetic DNA. It is also possible for the nucleic acid construct to be artificial nucleic acid, for example phosphorothioate RNA or DNA. Preferably the construct is DNA. Particularly preferred is when it is plasmid DNA.

The present invention further provides a host cell comprising an expression vector of the invention. Such cells include transient, or preferably stable higher eukaryotic cell lines, such as mammalian cells or insect cells, using for example a baculovirus expression system, lower eukaryotic cells, such as yeast or prokaryotic cells such as bacterial cells. Particular examples of cells which may be modified by insertion of vectors encoding for a polypeptide according to the invention include mammalian HEK293T, CHO, HeLa, NS0 and COS cells. Preferably the cell line selected will be one which is not only stable, but also allows for mature glycosylation of a polypeptide. Expression may be achieved in transformed oocytes. A polypeptide of the invention may be expressed in cells of a transgenic non-human animal, preferably a mouse or expressed into the milk of larger mammals, such as goats, sheep and cows. A transgenic non-human animal expressing a polypeptide of the invention is included within the scope of the invention. A polypeptide of the invention may also be expressed in *Xenopus laevis* oocytes.

The present invention also includes pharmaceutical or vaccine compositions, which comprise a therapeutically effective amount of polynucleotide or nucleic acid construct or polypeptide of the invention, optionally in combination with a pharmaceutically acceptable carrier, preferably in combination with a pharmaceutically acceptable excipient such as phosphate buffered saline (PBS), saline, dextrose, water, glycerol, ethanol, liposomes or combinations thereof. The vaccine composition may alternatively comprise a therapeutically effective amount of a nucleic acid construct of the invention, formulated onto metal beads, preferably gold beads. The vaccine composition of the invention may also comprise an adjuvant, such as, for example, in an embodiment, imiquimod, tucaresol or aluminium salts.

Preferably the adjuvant is administered at the same time as the immunogens of the present invention, and in preferred embodiments are formulated together. Such adjuvant agents contemplated by the invention include, but this list is by no means exhaustive and does not preclude other agents: synthetic imidazoquinolines such as imiquimod [S-26308, R-837], (Harrison, et al. 'Reduction of recurrent HSV disease using imiquimod alone or combined with a glycoprotein vaccine', *Vaccine* 19: 1820-1826, (2001)); and resiquimod [S-28463, R-848] (Vasilakos, et al. 'Adjuvant activities of immune response modifier R-848: Comparison with CpG ODN', *Cellular immunology* 204: 64-74 (2000).), Schiff bases of carbonyls and amines that are constitutively expressed on antigen presenting cell and T-cell surfaces, such as tucaresol (Rhodes, J. et al. 'Therapeutic potentiation of the immune system by costimulatory Schiff-base-forming drugs', *Nature* 377: 71-75 (1995)), cytokine, chemokine and co-stimulatory molecules, Th1 inducers such as interferon gamma, IL-2, IL-12, IL-15 and IL-18, Th2 inducers such as IL-4, IL-5, IL-6, IL-10 and other chemokine and co-stimulatory genes such as MCP-1, MIP-1 alpha, MIP-1 beta, RANTES, TCA-3, CD80, CD86 and CD40L, other immunostimulatory targeting ligands such as CTLA-4 and L-selectin, apoptosis stimulating proteins and peptides such as Fas, (49), synthetic lipid based adjuvants, such as vaxfectin, (Reyes et al., 'Vaxfectin enhances antigen specific antibody titres and maintains Th1 type immune responses to plasmid DNA immunization', *Vaccine* 19: 3778-3786) squalene, alpha-tocopherol, polysorbate 80, DOPC and cholesterol, endotoxin, [LPS], Beutler, B., 'Endotoxin, 'Toll-like receptor 4, and the afferent limb of innate immunity', *Current Opinion in Microbiology* 3: 23-30 (2000)) ; CpG oligo- and dinucleotides, Sato, Y. et al., 'Immunostimulatory DNA sequences necessary for effective intradermal gene immunization', *Science* 273 (5273): 352-354 (1996). Hemmi, H. et al., 'A Toll-like receptor recognizes bacterial DNA', *Nature* 408: 740-745, (2000) and other potential ligands that trigger Toll receptors to produce Th1-inducing cytokines, such as synthetic Mycobacterial lipoproteins, Mycobacterial protein p19, peptidoglycan, teichoic acid and lipid A.

Certain preferred adjuvants for eliciting a predominantly Th1-type response include, for example, a Lipid A derivative such as monophosphoryl lipid A, or preferably 3-de-O-acylated monophosphoryl lipid A. MPL[®] adjuvants are available from Corixa Corporation (Seattle, WA; see, for example, US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and

4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462. Immunostimulatory DNA sequences are also described, for
5 example, by Sato et al., *Science* 273:352, 1996. Another preferred adjuvant comprises a saponin, such as Quil A, or derivatives thereof, including QS21 and QS7 (Aquila Biopharmaceuticals Inc., Framingham, MA); Escin; Digitonin; or *Gypsophila* or *Chenopodium quinoa* saponins.

In particular, the adjuvant comprises an immunostimulatory CpG oligonucleotide,
10 such as disclosed in (WO96102555). Typical immunostimulatory oligonucleotides will be between 8-100 bases in length and comprises the general formula $X_1 \text{ CpGX}_2$ where X_1 and X_2 are nucleotide bases, and the C and G are unmethylated.

The preferred oligonucleotides for use in vaccines of the present invention preferably contain two or more dinucleotide CpG motifs preferably separated by at least three, more
15 preferably at least six or more nucleotides. The oligonucleotides of the present invention are typically deoxynucleotides. In a preferred embodiment the internucleotide in the oligonucleotide is phosphorodithioate, or more preferably a phosphorothioate bond, although phosphodiester and other internucleotide bonds are within the scope of the invention including oligonucleotides with mixed internucleotide linkages. e.g. mixed
20 phosphorothioate/phosphodiesters. Other internucleotide bonds which stabilise the oligonucleotide may be used. Methods for producing phosphorothioate oligonucleotides or phosphorodithioate are described in US5,666,153, US5,278,302 and WO95/26204.

Examples of preferred oligonucleotides have the following sequences. The sequences preferably contain phosphorothioate modified internucleotide linkages.

25 OLIGO 1: TCC ATG ACG TTC CTG ACG TT (CpG 1826)

OLIGO 2: TCT CCC AGC GTG CGC CAT (CpG 1758)

OLIGO 3: ACC GAT GAC GTC GCC GGT GAC GGC ACC ACG

OLIGO 4: TCG TCG TTT TGT CGT TTT GTC GTT (CpG 2006)

OLIGO 5: TCC ATG ACG TTC CTG ATG CT (CpG 1668)

30 Alternative CpG oligonucleotides may comprise the preferred sequences above in that they have inconsequential deletions or additions thereto.

The CpG oligonucleotides utilised in the present invention may be synthesized by any method known in the art (eg EP 468520). Conveniently, such oligonucleotides may be synthesized utilising an automated synthesizer. An adjuvant formulation for use in mice and containing CpG oligonucleotide can be purchased from Qiagen under the trade name "ImmunEasy". Preferably the adjuvant is one of the CpG's defines as OLIGO's 1, 2, 3, 4 or 5 adsorbed to aluminium hydroxide at an approximate 1:1 ratio weight/weight. OLIGO 4 is preferred for use in humans.

Preferably the CpG is in combination with a saponin, such as QS21, as described in WO 00/62800 and WO 00/09159 the contents of both of which is incorporated herein by reference.

Methods of treatment

The present invention provides novel treatments for atopic diseases, comprising an immunogen that is capable of generating an immune response in a vaccinee against IL-13. Most notably the present invention provides a method of treating an individual suffering from or being susceptible to COPD, asthma or atopic dermatitis, comprising administering to that individual a vaccine according to the present invention, and thereby raising in that individual a serum neutralising anti-IL-13 immune response and thereby ameliorating or abrogating the symptoms of COPD, asthma or atopic dermatitis.

Also provided by the present invention is the use of the immunogens of the present invention in the manufacture of a medicament for the treatment asthma. Also provided is a method of treatment of asthma comprising the administration to an individual in need thereof of a pharmaceutical composition or vaccine as described herein.

Preferably the pharmaceutical composition is a vaccine that raises an immune response against IL-13. The immune response raised is preferably an antibody response, most preferably an IL-13 neutralising antibody response.

The invention also provides:

- an expression vector which comprises a polynucleotide of the invention and which is capable of expressing a polypeptide of the invention;
- a host cell comprising an expression vector of the invention;

a method of producing a polypeptide of the invention which method comprises maintaining a host cell of the invention under conditions suitable for obtaining expression of the polypeptide and isolating the said polypeptide:

a vaccine composition comprising a polypeptide or polynucleotide of the invention and a
5 pharmaceutically acceptable carrier.

The methods of treatment of the present invention provide a method of treatment of asthma comprising one or more of the following clinical effects:

1. A reduction in airway hyper-responsiveness (AHR)
2. A reduction in mucus hyper-secretion and goblet cell metaplasia
- 10 3. A reduction in sub-epithelial fibrosis of the airways
4. A reduction in eosinophil levels
5. A reduction in the requirement for the use of inhaled corticosteroids (ICS) would also be a feature of successful treatment using an IL13 autovaccine.

15 The compositions of the present invention may be used for both prophylaxis and therapy. The present invention provides a polypeptide or a polynucleotide according to the invention for use in medicine. The invention further provides the use of a polypeptide or a polynucleotide of the invention in the manufacture of a medicament for the treatment of allergies, respiratory ailments such as asthma and COPD, helminth-infection related
20 disorders, fibrosis or cirrhosis of the liver.

The present invention also provides a method of vaccinating which comprises administering an effective amount of a vaccine composition of the invention to a patient and provoking an immune response to the vaccine composition.

The present invention also provides vaccine compositions as described herein for use
25 in vaccination of a mammal against IL-13 mediated disorders such as allergies, respiratory ailments, helminth-infection related disorders, fibrosis and cirrhosis of the liver. A vaccine composition capable of directing a neutralising response to IL-13 would therefore constitute a useful therapeutic for the treatment of asthma, particularly allergic asthma, in humans. It would also have application in the treatment of certain helminth infection-related disorders
30 (Brombacher, 2000 *Bioessays* 22:646-656) and diseases where IL-13 production is

implicated in fibrosis (Chiaramonte et al, 1999, *J Clin Inv* 104:777-785), such as chronic obstructive pulmonary disease (COPD) and cirrhosis of the liver.

The methods of treatment of the present invention provide a method of treatment of atopic dermatitis comprising one or more of the following clinical effects:

- 5 1. A reduction in skin irritation
2. A reduction in itching and scratching
3. A reduction in the requirement for conventional treatment.
4. if applicable a reduction in the requirement for the use of topical corticosteroids. An ideal IL13 autovaccine could potentially make ICS steroid treatment redundant, although a
10 reduction in the 'frequency of use' or 'dose required' of ICS is also envisaged as a valuable outcome.

The present invention also provides methods of treating or preventing IL-13 mediated disease, any symptoms or diseases associated therewith, comprising administering an effective amount of a protein, a polynucleotide, a vector or a pharmaceutical composition
15 according to the invention. Administration of a pharmaceutical composition may take the form of one or more individual doses, for example in a "prime-boost" therapeutic vaccination regime. In certain cases the "prime" vaccination may be via particle mediated DNA delivery of a polynucleotide according to the present invention, preferably incorporated into a plasmid-derived vector and the "boost" by administration of a recombinant viral vector
20 comprising the same polynucleotide sequence, or boosting with the protein in adjuvant. Conversely the priming may be with the viral vector or with a protein formulation typically a protein formulated in adjuvant and the boost with a DNA vaccine of the present invention.

The present invention provides methods of generating an anti self IL-13 antibody response in a host by the administration of vaccines of the present invention.

- 25 The vaccine compositions of the invention may be administered in a variety of manners for example via the mucosal, such as oral and nasal; pulmonary, intramuscular, subcutaneous or intradermal routes. Where the antigen is to be administered as a protein based vaccine, the vaccine will typically be formulated with an adjuvant and may be lyophilised and resuspended in water for injection prior to use. Such compositions may be
30 administered to an individual as an injectable composition, for example as a sterile aqueous

dispersion, preferably isotonic. Typically such compositions will be administered intramuscularly, but other routes of administration are possible.

One technique for intradermally administration involves particle bombardment (which is also known as 'gene gun' technology and is described in US Patent No. 5371015).

5 Proteins may be formulated with sugars to form small particles or DNA encoding the antigen may be coated on to inert particles (such as gold beads) and are accelerated at speeds sufficient to enable them to penetrate a surface of a recipient (e.g. skin), for example by means of discharge under high pressure from a projecting device. (Particles coated with nucleic acid vaccine constructs of the invention and protein sugar particles are within the
10 scope of the present invention, as are devices loaded with such particles.) Other methods of administering the nucleic acid constructs or compositions containing said constructs directly to a recipient include ultrasound, electrical stimulation, electroporation and microseeding which is described in US-5,697,901.

A nucleic acid construct of the present invention may also be administered by means
15 of specialised delivery vectors useful in gene therapy. Gene therapy approaches are discussed for example by Verme *et al*, Nature 1997, 389:239-242. Both viral and non-viral systems can be used. Viral based systems include retroviral, lentiviral, adenoviral, adeno-associated viral, herpes viral and vaccinia-viral based systems. Non-viral based systems include direct administration of nucleic acids and liposome-based systems. For example, the vectors may
20 be encapsulated by liposomes or within polylactide co-glycolide (PLG) particles.

A nucleic acid construct of the present invention may also be administered by means of transformed host cells. Such cells include cells harvested from a subject. The nucleic acid vaccine construct can be introduced into such cells *in vitro* and the transformed cells can later be returned to the subject. The nucleic acid construct of the invention may integrate into
25 nucleic acid already present in a cell by homologous recombination events. A transformed cell may, if desired, be grown up *in vitro* and one or more of the resultant cells may be used in the present invention. Cells can be provided at an appropriate site in a patient by known surgical or microsurgical techniques (e.g. grafting, micro-injection, etc.). Suitable cells include dendritic cells.

30 The amount of vaccine composition which is delivered will vary significantly, depending upon the species and weight of mammal being immunised, the nature of the

disease state being treated/protected against, the vaccination protocol adopted (i.e. single administration versus repeated doses), the route of administration and the potency and dose of the adjuvant compound chosen. Based upon these variables, a medical or veterinary practitioner will readily be able to determine the appropriate dosage level but it may be, for example, when the vaccine is a nucleic acid that the dose will be 0.5-5 μ g/kg of the nucleic acid constructs or composition containing them. In particular, the dose will vary depending on the route of administration. For example, when using intradermal administration on gold beads, the total dosage will preferably be between 1 μ g – 10ng, particularly preferably, the total dosage will be between 10 μ g and 1ng. When the nucleic acid construct is administered directly, the total dosage is generally higher, for example between 50 μ g and 1 or more milligram. The above dosages are exemplary of the average case.

In a protein vaccine, the amount of protein in each vaccine dose is selected as an amount which induces an immunoprotective response without significant, adverse side effects in typical vaccinees. Such amount will vary depending upon which specific immunogen is employed and how it is presented. Generally, it is expected that each dose will comprise 1-1000 μ g of protein, preferably 1-500 μ g, preferably 1-100 μ g, most preferably 1 to 50 μ g. An optimal amount for a particular vaccine can be ascertained by standard studies involving observation of appropriate immune responses in vaccinated subjects. Following an initial vaccination, subjects may receive one or several booster immunisation adequately spaced. Such a vaccine formulation may be either a priming or boosting vaccination regime; be administered systemically, for example *via* the transdermal, subcutaneous or intramuscular routes or applied to a mucosal surface *via*, for example, intra nasal or oral routes.

There can, of course, be individual instances where higher or lower dosage ranges are merited, and such are within the scope of this invention.

It is possible for the vaccine composition to be administered on a once off basis or to be administered repeatedly, for example, between 1 and 7 times, preferably between 1 and 4 times, at intervals between about 1 day and about 18 months, preferably one month. This may be optionally followed by dosing at regular intervals of between 1 and 12 months for a period up to the remainder of the patient's life. In an embodiment the patient will receive the antigen in different forms in a prime boost regime. Thus for example an antigen will be first

administered as a DNA based vaccine and then subsequently administered as a protein adjuvant base formulation. Once again, however, this treatment regime will be significantly varied depending upon the size and species of animal concerned, the amount of nucleic acid vaccine and / or protein composition administered, the route of administration, the potency
5 and dose of any adjuvant compounds used and other factors which would be apparent to a skilled veterinary or medical practitioner.

Throughout this specification the words “comprise” and “include” or variations such as “comprising”, “comprises”, “including”, “includes” etc., are to be construed both inclusively, that is, use of these words will imply the possible inclusion of integers or
10 elements not specifically recited and also in the exclusionary sense in that the words could be read as “consisting”.

As described herein, the present invention relates isolated polypeptides and isolated polynucleotides. In the context of this invention the term “isolated” is intended to convey that the polypeptide or polynucleotide is not in its native state, insofar as it has been purified
15 at least to some extent or has been synthetically produced, for example by recombinant methods, or mechanical synthesis. The term “isolated” therefore includes the possibility of the polypeptides or polynucleotides being in combination with other biological or non-biological material, such as cells, suspensions of cells or cell fragments, proteins, peptides, expression vectors, organic or inorganic solvents, or other materials where appropriate, but
20 excludes the situation where the polynucleotide is in a state as found in nature.

The present invention is exemplified, but not limited to, the following examples.

Example 1, *Methodology*

25 **For the methods below the following nomenclature applies:**

1. The construct called mouse IL13 (mIL-13) with tetanus toxin p30 epitope inserted into the protein (substituted into the looped region between alpha helices C and D of mouse IL13) is referred to as mIL13p30CD.
2. The construct called mouse IL13 with p30 at the N-terminus, is referred to as mIL13p30.
- 30 3. The construct called new chimaeric IL13 design with p30 N-terminus, is referred to as cIL13new.

IL-13 subcloning/ modifications:

A gene (mIL13CD) encoding mIL-13 containing the p30 epitope from tetanus toxin inserted into the CD loop was prepared synthetically. The synthetic gene contains a 5' *Kpn*I restriction site and a 3' *Bam*HI restriction site. This fragment was then subcloned between the *Kpn*I and *Bam*HI restriction sites of pCDN which encodes DHFR (Aiyer et al, 1994). The resultant intermediate was subsequently modified by inserting an FC fusion. Site-directed insertional mutagenesis was used to precisely insert human IgG1 FC in frame with the 3' end coding sequence preceding the stop codon of IL-13 (Geisser et al 2001). This was performed in two steps 1. IgG1 FC was amplified from a cDNA template, pCDN-FC, using the following primer set, (Forward : 5'..CAACTGTTTCGCCACGGCCCC TTCCTGGAGGTCCTGTTTCGGTGGACCAGGATCCGAGCCCAAATCGGCCGAC...3' and Reverse: 5' ...CTAGGTAGTTGGTAACCGTTAACGG...3') in a PCR reaction catalyzed by KOD proof-reading polymerase (Novagen). 2. The resultant PCR product was gel purified and 250ng used as a targeting fragment in a site-directed mutagenesis reaction using the QuickChange kit (Stratagene) with 50ng mIL-13 CD-pCDN and 2.5 U PfuTurbo. The mutagenesis protocol consisted of 18 Cycles of 30s at 95°C, 30S at 55°C, and 16 minutes at 68°C . At the end of the mutagenesis protocol, the reaction was digested with 10U *Dpn*I to remove the original methylated wild-type template DNA. 1ul of the final digested reaction was used to transform 100ul Epicurian chemically competent *E. coli* cells (Stratagene). Recombinant clones were screened by restriction digestion and positive clones sequence confirmed fully across the FC region using IL-13 forward and pCDN reverse primers. The final plasmid, pCDNmIL13CDFC encodes a C-terminal FC fusion separated by a PreScission protease cleavage site for FC removal. Transcription is under control of the CMV₄ promoter.

The complete sequence of the insert is shown in Figure 18 (SEQ ID NO. 23).

pCDNmIL13p30FC was constructed in exactly the same way as described above for pCDNmIL13CDFC, replacing the mIL13CD synthetic gene with one where the p30 epitope was present at the N terminus of the mature protein instead of being in the CD loop. The same forward and reverse primers were used to generate the targeting fragment for site-

directed insertion of the FC region into pCDNmIL13p30. The complete sequence of the insert is shown in Figure 19 (SEQ ID NO. 24)

pCDNcIL13newFC was constructed using a synthetic gene encoding the cIL13new molecule and the following forward primer

5 (5'..AACCTGTTTCGCCGCGGCCCTTCCTGGAGGTCC
TGTTCCGGTGGACCAGGATCCGAGCCCAAATCGGCCGAC...3', (SEQ ID NO. 25)) and the same reverse primer described above to generate the targeting fragment for site-directed insertion of the FC region into pCDNcIL13new. The complete sequence of the insert is shown in Figure 20 (SEQ ID NO. 26)).

10 pCDN IL13oldFC was constructed by site-directed replacement of mIL13 CD within pCDNmIL13CDFC with mouse chimeric IL13 (see WO 02/070711). Site-directed replacement was performed as described for site-directed insertion. cIL13 was PCR amplified from 6His-cIL13 using the following primers (Forward: 5' 5'...GTGTCTCTCC CTCTGACCCTTAGG...3' (SEQ ID NO. 27) and Reverse:

15 5'...CAGTTGCTTTGTGTAGCTGAG CAG...3' (SEQ ID NO. 28) to generate a targeting fragment for replacement into pCDNmIL13. This generates a precise fusion to the IL-13 signal sequence encoded at the 5' end and the PreScission-FC region encoded at the 3' end. The complete sequence of the insert is shown in Figure 21 (SEQ ID NO. 29).

20 In all of Figures 18 to 21, doubly underlined amino acid residues indicate the secretion signal sequence (removed in the course of expression and secretion from the host cell), single underlined residues, the Precision protease site and italicised residues the Fc fusion partner.

Generation of Stable CHO E1A clones:

25 Plasmids were stably expressed in a DHFR negative, E1A expressing line (CHO E1A, ACC317). Cells were resuspended at 1×10^7 cell/ml in cold phosphate buffered sucrose, transferred to a Gene Pulser Cuvette, and electroporated with 15ug *Not* I linearized plasmid at 400volt and 25uF in a GenePulser (Biorad). Electroporated cells were plated in a 96 well plate at 2.5×10^3 viable cells per well in complete medium containing 1 X Nucleosides.

30 After 48 hours the medium was exchanged with fresh medium lacking nucleosides. Cells

were subsequently selected over 3-4 weeks in the absence of nucleosides. Positive clones were screened from the 96 well plate by monitoring FC expression from conditioned medium using an FC- electrochemiluminescence detection protocol (Yang, et al., 1994) on an Origen analyzer (IGEN). Positive cell lines were scaled to several litres in complete medium minus
5 nucleosides. Fermentations were carried out at 34°C for 10-11 days. Conditioned medium was harvested and 0.2 uM sterile filtered in preparation for FC purification.

Purification:

Murine IL13CD/Fc was captured from CHO medium onto ProSep-A High Capacity resin
10 (Bioprocessing Limited). The murine IL13CD/Fc was eluted from the ProSep-A resin with 0.1M Glycine pH=3.0, neutralized with 1M HEPES pH=7.6, and dialyzed against 25mM sodium phosphate 0.15M sodium chloride pH=7 (Spectra/Por® 7 membrane, MWCO:8000). Overall yield was 644mg murine IL13CD/Fc from 3.8 liter CHO medium. Other IL13/Fc fusion proteins were prepared similarly.

15 Before use in vaccination studies, the Fc portions of these molecules were cleaved off using Precision protease and removed. The resulting vaccine preparations comprise essentially those amino acid residues indicated in Figures 18 to 21 by plain text (ie neither underlined nor italicised).

20

References:

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30

Example 2, Efficacy of an anti-IL13 vaccine in a mouse asthma model.

The mouse asthma model.

The ovalbumin challenge mouse asthma model is routinely used to assess the efficacy of asthma therapeutic treatments *in vivo*. Mice are sensitised with 2 intra-peritoneal doses of ovalbumin given 7 days apart, which establishes the sensitivity of the mice to ovalbumin. The asthmatic phenotype can then be generated by giving 3 intra-nasal doses of ovalbumin. Mice subjected to this protocol exhibit a high level of airway hyper-responsiveness to the spasmogen 5HT, inflammation of the lung (most notably an eosinophilia of the lung tissue and broncho-alveolar lavage fluid), and a massive goblet cell metaplasia (and associated mucus hyper-secretion) of the lung airway epithelium. This phenotype mimics that seen in human asthmatics. (Similar mouse asthma models are described in Science 1998 vol 282, pp:2258 – 2261 and 2261 – 2263). This model is also described in WO 02/070711.

Anti-IL13 vaccine treatment.

Two anti-IL13 vaccine treatments were assessed for efficacy in the ovalbumin challenge mouse asthma model, in mice that had previously been sensitised to ovalbumin (Sigma UK Ltd, Poole, Dorset). Both are based on the mouse chimeric IL13 molecule disclosed in WO 02/070711, which is expressed and purified as a fusion protein with GST. It is here referred to as gst-cIL13.

1. Vaccine 1 = gst-cIL13 + 'ImmunEasy' adjuvant (Qiagen, Cat.No. 303101)
2. Vaccine 2 = gst-cIL13 + liposomes comprising cholesterol in combination with 10 µg 3-de-O-acylated monophosphoryl lipid A (3D-MPL) and 10µg QS21 saponin (see EP0822831B1, SmithKline Beecham Biologicals S.A.)

Negative control vaccine treatment groups were also included.

3. Negative control for vaccine 1 = gst + 'ImmunEasy' adjuvant
4. Negative control for vaccine 2 = gst + liposomes comprising cholesterol in combination with 10 µg 3-de-O-acylated monophosphoryl lipid A (3D-MPL) and 10µg QS21 saponin (see EP0822831B1, SmithKline Beecham Biologicals S.A.).

Following sensitisation with ovalbumin mice were immunised with 4 doses of vaccine, each vaccine dose given 4 weeks apart over a 12 week period. Mice were then challenged with ovalbumin and the asthmatic phenotype assessed.

Other control treatment groups in the efficacy study.

A. Dexamethasone (Sigma UK Ltd, Poole, Dorset) is a gold-standard steroid treatment routinely used in this mouse asthma model. Mice were given 3 doses of 1.5mg/kg dexamethasone via the intra-peritoneal route, during ovalbumin challenge.

5 B. Passively administered anti-mouse IL13 polyclonal antibody (a protein A purified reagent previously made in-house in rabbits) was given as a positive control treatment in this mouse asthma model. A dose of antibody previously shown to generate full anti-IL13 driven efficacy in this mouse asthma model was administered during ovalbumin challenge (= 3 doses of 0.5ml of a stock having an endpoint titre of 2×10^5 , for further details see WO
10 02/070711 A1)

C. The maximum phenotype generated by this model was established in a negative control treatment group using saline (Fresenius Kabi, Warrington, UK). Mice were given 3 doses of saline by the intra-nasal route during ovalbumin challenge. Saline treatment shows no efficacy in this model, therefore the most severe asthmatic phenotype is generated.

15 D. As a baseline for comparison of the asthma model phenotype to 'no induced asthmatic phenotype', one treatment group was only sensitised with ovalbumin, no ovalbumin challenge doses were given. These mice exhibit normal lung physiology.

**Serum IL13 neutralisation capacity generated in mice immunised with the anti-IL13
20 vaccines, or passively administered anti-IL13 polyclonal antibody.**

At the end of the mouse asthma model, mice treated with vaccine or passively administered anti-IL13 polyclonal antibody, had serum samples analysed for IL13 neutralisation capacity using the mouse IL13-induced TF-1 cell proliferation assay, as described in WO 02/070711. This analysis yields a neutralisation measure termed ND_{50} ,
25 which represents the maximum dilution of mouse serum which is able to reduce by 50% the bioactivity of 5ng/ml of mouse IL13 in a TF-1 cell proliferation assay.

Our previous data also demonstrated that, using passively administered neutralising anti-IL13 antibodies, maximal efficacy in this murine asthma model is correlated with a serum ND_{50} value of approximately 1/476. This critical level of neutralisation we term ED_{100}
30 (the effective neutralising dose required to give 100% efficacy), and commonly express serum neutralisation capacities relative to this level. For example, a serum sample which had

a ND₅₀ of 1/952 would be said to have a neutralising capacity of 2.0 x ED₁₀₀. A sample with a ND₅₀ of 1/238 would have a neutralisation capacity of 0.5 x ED₁₀₀.

The serum IL13 neutralisation capacity data from this experiment are shown in Figure 22, and are plotted as a multiples of ED₁₀₀.

5 All mice that were treated with the chimeric IL13 vaccine or passively administered with anti-IL13 polyclonal antibody generated serum neutralisation in excess of 1 x ED₁₀₀. Therefore it was predicted that the mice in these treatment groups would receive full anti-IL13 driven benefit in the asthma model.

Airways hyper-responsiveness (AHR) data.

10 Dose response curves to inhaled spasmogens are used to determine the response of the airways to a bronchoconstrictor stimulus. These curves are comprised of two main components:

1. Hypersensitivity - a leftward shift in the dose response curve (DRC)
2. Hyperreactivity - an increase slope of the DRC and/or a loss in the plateau response

15 These components together give rise to the general term 'bronchial or airway hyperresponsiveness' (BHR or AHR) and this is typically defined as '*an increase in the ease and degree of airway narrowing in response to bronchoconstrictor stimuli*'.

AHR was measured by challenging conscious mice with a dose of 5HT spasmogen, and then measuring the effects on respiratory flow and volume parameters using a whole-
20 body plethysmography apparatus (Buxco, Sharon, CT). The preferred readout parameter from this analysis is the measure of enhanced pause (PENH). Figure 23 illustrates AHR data from this experiment obtained by plotting PENH area under curve values for a 5HT spasmogen concentration of 3mg/ml. Data points are the means and standard errors for the treatment groups indicated.

25

Both the vaccine treatments and passively administered anti-IL13 polyclonal antibody were as effective as dexamethasone at reducing the level of AHR. The negative control vaccine treatments did not reduce AHR.

Lung inflammation data.

Lung inflammatory cell content was assessed in the broncho-alveolar lavage fluid (BAL). Average numbers of eosinophils, macrophages, lymphocytes and neutrophils were plotted against treatment received (Figure 24).

Both the vaccine treatments and passively administered anti-IL13 polyclonal antibody were as effective as dexamethasone at reducing the level of eosinophils in the BAL fluid. Interestingly, the negative control treatment gst + 'ImmunEasy' also appeared to effectively reduce the level of BAL eosinophilia. This is probably due to the activity of the CpG component in the 'ImmunEasy' adjuvant which is known to be an immunomodulatory compound with pro-Th1 activity.

10 **Goblet cell metaplasia and mucus hyper-secretion data.**

Mucus containing goblet cells are not normally present at significant frequencies in the mouse airway epithelium. Following sensitisation and challenge with ovalubumin in this asthma model, the airway epithelium becomes densely packed with mucus containing goblet cells due to a metaplasia of the epithelial layer.

15 Following fixation, representative samples of the lungs from each animal were processed for paraffin histology. Sections were cut at 5 μ and stained with ABPAS (Alcian blue periodic acid Schiff's reagent, BDH-Merck) with α -amylase (Sigma UK Ltd, Poole, Dorset) pre-digestion for histopathological evaluation of airway goblet cells (preparative histology by ProPath UK Ltd, Hereford, UK).

20 The lung sections stained with ABPAS were scored for goblet cell numbers using the 6-point semi-quantitative scoring system shown below. The results are shown in Figure 25..

SCORING SYSTEM FOR GOBLET CELLS

Score	Observation
25 0	No goblet cells
1	Very few goblet cells
2	Low numbers of goblet cells
3	Moderate numbers of goblet cells
4	Heavy numbers of goblet cells
30 5	Massive numbers of goblet cells

Note that the scoring system is not linear, and that the difference between a score of 2 or 3 is highly significant in relation to the number of goblet cells present in the epithelium.

Representative sections for some of the treatment groups are shown in Figure 26A, gst-cIL13 + 'ImmunEasy'; Figure 26B, gst-'ImmunEasy'; Figure 27A, gst-cIL13 +
5 Liposomes comprising cholesterol in combination with 10 µg 3-de-O-acylated monophosphoryl lipid A (3D-MPL) and 10µg QS21 saponin (see EP0822831B1, SmithKline Beecham Biologicals S.A.); Figure 27B, gst + Liposomes comprising cholesterol in combination with 10 µg 3-de-O-acylated monophosphoryl lipid A (3D-MPL) and 10µg QS21 saponin (see EP0822831B1, SmithKline Beecham Biologicals S.A.); Figure 28,
10 dexamethasone; Figure 29, maximum asthmatic phenotype.

Both the vaccine treatments and passively administered anti-IL13 polyclonal antibody dramatically reduced the numbers of mucus-containing goblet cells in the airway epithelium. The reduction in goblet cell number is highly significant for all anti-IL13 treatments versus the saline (maximum phenotype) treatment group ($p < 0.01$). Negative
15 control vaccines had no effect. Dexamethasone treatment had very little effect on goblet cell metaplasia (GCM) in this study.

Summary.

The anti-IL13 vaccine treatments were very effective at abrogating the asthmatic
20 phenotype in the mouse asthma model. Anti-IL13 vaccine was as effective as dexamethasone for treatment of AHR and eosinophilia, and was superior to dexamethasone for treatment of goblet cell metaplasia and mucus hyper-secretion.

Example 3, Correlation of goblet cell metaplasia with the level of serum IL13 neutralisation
25 *capacity.*

Some animals immunised with the anti-IL13 vaccines achieved serum IL13 neutralisation levels of less than $1.0 \times ED_{100}$. To determine whether these animals were receiving any discernible benefit (keeping in mind that ED_{100} is defined in terms of maximal benefit), they too were challenged with ovalbumin, and the degree of GCM determined.
30 The data below indicates the relationship between goblet cell metaplasia score and level of IL13 neutralisation capacity induced in the serum by the vaccine.

SCORING SYSTEM FOR GOBLET CELLS

<i>Score</i>	<i>Observation</i>
0	No goblet cells
5 1	Very few goblet cells
2	Low numbers of goblet cells
3	Moderate numbers of goblet cells
4	Large numbers of goblet cells
5	Massive numbers of goblet cells

10

The Goblet cell data is shown in table 1 below and in Figure 30:

Table 1,

	GCM	neut.
Mouse	score	capacity
A1	2.5	0.41
2	3	0.3
4	3.5	0.31
8	3.5	0.21
9	3.5	0
10	2	0.8
11	1.5	0.36
12	3	0.37
14	3	0
15	2.5	0.3
16	2.5	0.34
18	3	0
20	3.5	0
C30	3	0
31	3	0.21
33	3	0
34	4	0
35	3.5	0
36	3	0
38	3	0.24
41	2.5	0.36
42	3	0.34
43	3.5	0
45	3	0
46	1.5	0.8
47	2.5	0.31
48	2	0.26

Only mice that generated serum IL13 neutralisation capacity less than 1 x ED₁₀₀ were included in this analysis, because, by definition, animals with a serum IL13 capacity equal to 5 or in excess of 1 x ED₁₀₀ achieve a maximal efficacy in respect of suppressing goblet cell metaplasia.

The data indicates that there is a correlation between the level of serum IL13 neutralisation capacity and the severity of goblet cell metaplasia ($R^2 = 0.52$). The higher the level of IL13 neutralisation, the lower the severity of goblet cell metaplasia.

10 These data, together with those of Example 3, validate the use of the ED100 measure as a powerful predictor of efficacy of anti-IL13 treatments against the asthmatic phenotype.

Any vaccine, antibody, soluble receptor or other IL13 neutralising treatment may be evaluated as follows:

1. Administer the IL13 neutralising treatment to the recipient at the desired dose and frequency.
- 5 2. Take a serum sample.
3. Determine the IL13 ND₅₀ of the serum sample by analysing it, and dilutions thereof, in a IL13 bioassay such as the TF1 proliferation assay. The bioassay is chosen such that it is possible to determine the greatest serum dilution which causes a 50% inhibition of the specific effect of 5 ng/ml of mouse IL13. For treatments directed to human IL13, the
10 TF1 bioassay may still be used, but the stimulating cytokine will be human IL13 used at a concentration in the range 3-6 ng/ml.
4. Divide the ND₅₀ value obtained by 1/476 to produce a ED₁₀₀ multiple.
5. If this multiple is 1.0 or greater, the IL13 neutralising treatment is expected to have maximal efficacy on the asthmatic phenotype.
- 15 6. If the multiple is considerably less than 1.0, for example 0.2 or less, then no significant efficacy is to be expected.
7. If the multiple lies between these limits, then some efficacy may be seen, but it will not be optimal, indicating that improvements in the treatment will be desirable.

This process may be used to guide dose selection for maximal efficacy. If, after an initial
20 number of doses of agent, the serum IL13 neutralisation capacity has not reached a level at least equal to 1.0 x ED₁₀₀, then further doses are given to bring the neutralisation capacity up to this level.

Example 4, *Immunogenicity of an anti-IL13 protein vaccine in combination with various*
25 *adjuvants.*

Studies to investigate the immunogenicity of a gst-cIL-13 immunogen, with or without the additional promiscuous T-cell epitope P30, in combination with several different adjuvants were performed.

gst-cIL13 protein immunogenicity studies

30 BalbC mice were immunised with 100µg gst-cIL13 in adjuvant for the primary immunisation, followed by 50µg gst-cIL13 in adjuvant for the boost immunisations.

Immunisations were administered on a four weekly basis, serum samples taken from mice 2 weeks after each immunisation (to monitor the level of IL13 neutralisation capacity generated by these antibodies in the serum sample). The g_{st}-cIL-13 immunogen was combined with four different adjuvants:

- 5 Group A CpG-2006 adsorbed onto aluminium hydroxide
 Group B CpG-1826
 Group C CFA prime/IFA boost
 Group D aluminium hydroxide

CpG-2006 and CpG-1826 are oligonucleotides containing unmethylated CG
 10 dinucleotides, and well-known in the literature for possessing immunostimulatory activity. CFA/IFA denote complete and incomplete Freund's adjuvant respectively.

The IL13 neutralisation capacity generated by these antibodies in serum samples was measured in a mouse IL13 bioassay (the TF-1 cell proliferation assay). The table below shows the results (expressed as a multiple of ED₁₀₀) for day 99, post 4 immunisations. The
 15 data is also represented graphically in Figure 31. In this figure, and in the similar figures that follow, each dot indicates a serum IL13 neutralisation measurement for one animal. Animals whose serum neutralising capacity is below the sensitivity threshold of the assay (<0.2 x ED₁₀₀) are not plotted.

**IL13 neutralisation capacity
 expressed as ED₁₀₀**

	Adjuvant treatment			
	A	B	C	D
1	<0.2	<0.2	<0.2	<0.2
2	2.7	<0.2	<0.2	<0.2
3	0.5	<0.2	<0.2	<0.2
4	<0.2	1.4	<0.2	<0.2
5	<0.2	<0.2	<0.2	<0.2

Adjuvant A (CpG (2006) adsorbed onto aluminium hydroxide), in combination with gst-cIL13 protein, was the most effective at generating neutralising anti-IL13 antibody responses. No neutralising anti-IL13 antibody responses were detected for mice treated with gst-cIL13 protein combined with either alum or CFA/IFA adjuvants.

5

p30-cIL13 protein.

Study 1

For this study a different form of IL13 vaccine was used. This is another chimeric IL13 molecule which contains the p30 epitope from tetanus toxin at the N terminus. It is encoded by the plasmid pCDNcIL13newFC (Figure 20), and prepared for vaccine studies as described in Example 1. The fully processed molecule is termed p30-cIL13 in the descriptions below.

Five CD-1 mice were immunised with 40µg p30-cIL13 in adjuvant for the primary immunisation, followed by 40µg p30-cIL13 in adjuvant for the boost immunisations. Immunisations were administered on a four weekly basis, serum samples taken from mice 2 weeks after each immunisation (to monitor the level of anti-mouse IL13 antibodies present, and the IL13 neutralisation capacity generated by these antibodies in the serum sample). As a negative control, serum samples were also analysed from three unimmunised CD-1 mice.

Group	Adjuvant
A	Immuneasy™ (purchased from Qiagen Corp.)
B	liposomes comprising cholesterol in combination with 10 µg 3-de-O-acylated monophosphoryl lipid A (3D-MPL) and 10µg QS21 saponin (see EP0822831B1, SmithKline Beecham Biologicals S.A.).
C	No immunisations

25

Anti-mouse IL13 antibody levels (in a 1/100 dilution of the serum samples) were measured by ELISA. The table below shows the results (expressed as absorbance at 490nm) for day 63 post 3 immunisations. The data is also represented graphically in Figure 32, where each bar represents the data for a single mouse.

30

ELISA data

Absorbance @ 490nm

	Mouse				
	1	2	3	4	5
A	2.654	2.377	2.0995	1.5925	2.4125
B	2.81	2.398	n/a	2.6775	2.95
C	0.049	0.0595	0.1095		

(n/a = sample not available)

Both adjuvants combined with p30-cIL13 protein were able to raise anti-IL13 antibody responses in CD-1 mice.

The IL13 neutralisation capacity generated by these antibodies in serum samples was measured in a mouse IL13 bioassay (the TF-1 cell proliferation assay). The table below shows the results (expressed as a multiple of ED₁₀₀) for day 63, post 3 immunisations. The data is also represented graphically in Figure 33.

IL13 neutralisation capacity expressed as ED ₁₀₀		
CD-1 mice	A	B
1	0.755	4.444
2	<0.2	2.963
3	<0.2	n/a
4	<0.2	11.429
5	<0.2	3.077

Adjuvant B, in combination with p30-cIL13 protein, was the most effective at generating neutralising anti-IL13 antibody responses, 4 out of 5 mice generating potent anti-IL13 neutralising antibody responses in excess of 1 x ED₁₀₀. In comparison, only 1 mouse generated neutralising anti-IL13 antibody responses when treated with p30-cIL13 protein combined with ImmunEasy adjuvant (adjuvant A).

Study 2

p30-cIL13 protein with oil emulsion adjuvant with 3D-MPL and QS21.

Five CD-1 mice were immunised with 40µg p30-cIL13 in adjuvant for the primary immunisation, followed by 40µg p30-cIL13 in adjuvant for the boost immunisations. Immunisations were administered on a four weekly basis, serum samples taken from mice 2 weeks after each immunisation (to monitor the level of anti-mouse IL13 antibodies present, and the IL13 neutralisation capacity generated by these antibodies in the serum sample). As a negative control, serum samples were also analysed from three unimmunised CD-1 mice.

Group	Adjuvant
A	ImmunEasy™
B	oil in water emulsion (oil phase: 1:1 v/v squalene:alpha tocopherol mix, cholesterol + TWEEN 80™ surfactant) + 10µg 3D-MPL and 10µg QS21) (for further details see WO 99/11241 (described as SB62c'))
C	no immunisations

Anti-mouse IL13 antibody levels (in a 1/100 dilution of the serum samples) were measured by ELISA. The table below shows the results (expressed as absorbance at 490nm) for day 63 post 3 immunisations. The data is also represented graphically in Figure 34.

ELISA data	Absorbance @ 490nm				
	Mouse				
	1	2	3	4	5
A	2.654	2.377	2.0995	1.5925	2.4125
B	2.8165	2.906	2.9035	n/a	3.081
C	0.049	0.0595	0.1095		

Both adjuvants combined with p30-cIL13 protein were able to raise anti-IL13 antibody responses in CD-1 mice.

The IL13 neutralisation capacity generated by these antibodies in serum samples was measured in a mouse IL13 bioassay (the TF-1 cell proliferation assay). The table below shows the results (expressed as a multiple of ED₁₀₀) for day 63, post 3 immunisations. The data is also represented graphically in Figure 35.

**IL13 neutralisation capacity
expressed as ED₁₀₀**

CD-1	A	B
mice		
1	0.755	3.077
2	<0.2	9.524
3	<0.2	3.333
4	<0.2	n/a
5	<0.2	1.176

Adjuvant B, in combination with p30-cIL13 protein, was the most effective at generating neutralising anti-IL13 antibody responses, 4 out of 5 mice generating potent anti-IL13 neutralising antibody responses in excess of 1 x ED₁₀₀. In comparison, only 1 mouse
 5 generated neutralising anti-IL13 antibody responses when treated with p30-cIL13 protein combined with ImmunEasy adjuvant (group A).

Study 3

p30-cIL13 protein with oil emulsion adjuvant (without immunostimulant).

10 Five CD-1 mice were immunised with 40µg p30-cIL13 in adjuvant for the primary immunisation, followed by 40µg p30-cIL13 in adjuvant for the boost immunisations. Immunisations were administered on a four weekly basis, serum samples taken from mice 2 weeks after each immunisation (to monitor the level of anti-mouse IL13 antibodies present, and the IL13 neutralisation capacity generated by these antibodies in the serum sample). As
 15 a negative control, serum samples were also analysed from three unimmunised CD-1 mice.

Group	Adjuvant
A	ImmunEasy™
B	oil in water emulsion (oil phase: 1:1 v/v squalene:alpha tocopherol
20	mix, cholesterol + TWEEN 80™ surfactant) (for details see WO9517210)
C	no immunisations

Anti-mouse IL13 antibody levels (in a 1/100 dilution of the serum samples) were measured by ELISA. The table below shows the results (expressed as absorbance at 490nm) for day 63 post 3 immunisations. The data is also represented graphically in Figure 36, where each bar represents the data for a single mouse.

5

ELISA data	Absorbance @ 490nm				
	Mouse				
	1	2	3	4	5
A	2.654	2.377	2.0995	1.5925	2.4125
B	n/a	3.038	1.5625	n/a	n/a
C	0.049	0.0595	0.1095		

Both adjuvants combined with p30-cIL13 protein were able to raise anti-IL13 antibody responses in CD-1 mice.

The IL13 neutralisation capacity generated by these antibodies in serum samples was measured in a mouse IL13 bioassay (the TF-1 cell proliferation assay). The table below shows the results (expressed as a multiple of ED₁₀₀) for day 63, post 3 immunisations. The data is also represented graphically in Figure 37.

IL13 neutralisation capacity expressed as ED ₁₀₀		
CD-1 mice	A	B
1	0.755	n/a
2	<0.2	0.32
3	<0.2	0.69
4	<0.2	n/a
5	<0.2	n/a

Adjuvant B, in combination with p30-cIL13 protein, was the most effective at generating neutralising anti-IL13 antibody responses, 2 out of 5 mice generating anti-IL13

neutralising antibody responses. In comparison, only 1 mouse generated neutralising anti-IL13 antibody responses when treated with p30-cIL13 protein combined with ImmunEasy adjuvant (adjuvant A).

5 *Summary*

The ability of the P30 immunogens to augment the immune response in the outbred CD-1 mouse strain is significant in that it suggests that the use of this immunogen is not limited to a single immunological background, and the advantageous effects of P30 should also be obtained in an outbred human clinical setting.